#### Appendix I

### Version With Markings To Show Changes Made in accordance with 37 C.F.R. § 1.121(c)(1)(ii)

#### In The Claims:

Please cancel Claims 36-41 and 43-60.

Please amend the following Claims:

- 26. A method for detecting the presence of a target nucleic acid molecule [by detecting non-target cleavage products] comprising:
  - a) forming a cleavage structure comprising: [providing:
    - i) a cleavage agent;]
  - [i]i) a synthetic target nucleic acid, said synthetic target nucleic acid comprising a first region and a second region, said second region downstream of and contiguous to said first region;
  - [i]ii) a first <u>nucleic acid molecule</u> [oligonucleotide], wherein at least a portion of said first <u>nucleic acid molecule</u> [oligonucleotide] is completely complementary to said first portion of said first target nucleic acid;
  - iii[v]) a second <u>nucleic acid molecule</u> [oligonucleotide] comprising a 3' portion and a 5' portion, wherein said 5' portion is completely complementary to said second portion of said target nucleic acid;
  - b) <u>cleaving said cleavage structure with a thermostable 5' nuclease so as to generate non-target cleavage product; and [mixing said cleavage agent, said synthetic target nucleic acid, said first oligonucleotide and said second oligonucleotide to create a reaction mixture under reaction conditions such that at least said portion of said first oligonucleotide is annealed to said first region of said target nucleic acid and wherein at least said 5' portion of said second oligonucleotide is annealed to said second region of said target nucleic acid so as to create a cleavage structure, and wherein cleavage of said cleavage structure occurs to generate non-target cleavage product; and]</u>
    - c) detecting the cleavage of said cleavage structure.

Please add the following claims:

- 61. The method of Claim 26, wherein said cleaving step is conducted under isothermal conditions.
- 62. The method of Claim 26, wherein said thermostable 5' nuclease comprises a 5' nuclease of a DNA polymerase.
- 63. The method of Claim 62, wherein said DNA polymerase is *Taq* DNA polymerase.
- 64. The method of Claim 26, wherein said 3' portion of said second nucleic acid molecule comprises an aromatic ring.
- 65. The method of Claim 26, wherein said 3' portion of said second nucleic acid molecule comprises a 3' terminal nucleotide not complementary to said target nucleic acid.
- 66. The method of Claim 26, wherein said 3' portion of said second nucleic acid molecule consists of a single nucleotide.
- 67. The method of Claim 66, wherein said single nucleotide is not complementary to said target nucleic acid.
- 68. The method of Claim 66, wherein said single nucleotide is complementary to said target nucleic acid.
- 69. The method of Claim 65, wherein said 3' terminal nucleotide comprises a naturally occurring nucleotide.

- 70. The method of Claim 65, wherein said 3' terminal nucleotide comprises a nucleotide analog.
- 71. The method of Claim 26, wherein a plurality of said first nucleic acid molecule is provided, such that said first nucleic acid molecule is in concentration excess compared to said target nucleic acid.
- 72. The method of Claim 26, wherein a plurality of said second nucleic acid molecule is provided, such that said second nucleic acid molecule is in concentration excess compared to said target nucleic acid.
- 73. The method of Claim 26, wherein said target nucleic acid and said second nucleic acid form a duplex, and wherein a plurality of said first nucleic acid is provided such that said first nucleic acid molecule is in concentration excess compared to said duplex.
- 74. The method of Claim 73, wherein said cleaving said cleavage structure comprises cleaving said first nucleic acid molecule to generate non-target cleavage product.
- 75. The method of Claim 74, wherein said non-target cleavage product from said first nucleic acid molecule is generated in concentration excess compared to said duplex.
- 76. The method of Claim 26, further comprising providing a third nucleic acid molecule complementary to a third portion of said target nucleic acid upstream of said first portion of said first target nucleic acid, wherein said cleavage structure comprises said third nucleic acid molecule.

#### Appendix II

## Entire Set Of Pending Claims pursuant to 37 C.F.R. § 1.121(c)(3)

- 26. (Amended) A method for detecting the presence of a target nucleic acid molecule comprising:
  - a) forming a cleavage structure comprising:
  - i) a synthetic target nucleic acid, said synthetic target nucleic acid comprising a first region and a second region, said second region downstream of and contiguous to said first region;
  - ii) a first nucleic acid molecule, wherein at least a portion of said first nucleic acid molecule is completely complementary to said first portion of said first target nucleic acid;
  - iii) a second nucleic acid molecule comprising a 3' portion and a 5' portion, wherein said 5' portion is completely complementary to said second portion of said target nucleic acid;
  - b) cleaving said cleavage structure with a thermostable 5' nuclease so as to generate non-target cleavage product; and
    - c) detecting the cleavage of said cleavage structure.
- 27. The method of Claim 26, wherein said detecting the cleavage of said cleavage structure comprises detecting said non-target cleavage product.
- 28. The method of Claim 26, wherein said synthetic target nucleic acid comprises an amplified nucleic acid.
- 29. The method of Claim 28, wherein said amplified nucleic acid is produced using a polymerase chain reaction.
- 30. The method of Claim 26, wherein said detecting the cleavage of said cleavage structure comprises detection of fluorescence.

- 31. The method of Claim 26, wherein said detecting the cleavage of said cleavage structure comprises detection of mass.
- 32. The method of Claim 26, wherein said detecting the cleavage of said cleavage structure comprises detection of fluorescence energy transfer.
- 33. The method of Claim 26, wherein said detecting the cleavage of said cleavage structure comprises detection selected from the group consisting of detection of radioactivity, luminescence, phosphorescence, fluorescence polarization, and charge.
- 34. The method of Claim 26, wherein said first oligonucleotide is attached to a solid support.
- 35. The method of Claim 26, wherein said second oligonucleotide is attached to a solid support.
- 42. The method of Claim 26, wherein said synthetic target nucleic acid comprises DNA.
- 61. The method of Claim 26, wherein said cleaving step is conducted under isothermal conditions.
- 62. The method of Claim 26, wherein said thermostable 5' nuclease comprises a 5' nuclease of a DNA polymerase.
- 63. The method of Claim 62, wherein said DNA polymerase is *Taq* DNA polymerase.
- 64. The method of Claim 26, wherein said 3' portion of said second nucleic acid molecule comprises an aromatic ring.

- 65. The method of Claim 26, wherein said 3' portion of said second nucleic acid molecule comprises a 3' terminal nucleotide not complementary to said target nucleic acid.
- 66. The method of Claim 26, wherein said 3' portion of said second nucleic acid molecule consists of a single nucleotide.
- 67. The method of Claim 66, wherein said single nucleotide is not complementary to said target nucleic acid.
- 68. The method of Claim 66, wherein said single nucleotide is complementary to said target nucleic acid.
- 69. The method of Claim 65, wherein said 3' terminal nucleotide comprises a naturally occurring nucleotide.
- 70. The method of Claim 65, wherein said 3' terminal nucleotide comprises a nucleotide analog.
- 71. The method of Claim 26, wherein a plurality of said first nucleic acid molecule is provided, such that said first nucleic acid molecule is in concentration excess compared to said target nucleic acid.
- 72. The method of Claim 26, wherein a plurality of said second nucleic acid molecule is provided, such that said second nucleic acid molecule is in concentration excess compared to said target nucleic acid.
- 73. The method of Claim 26, wherein said target nucleic acid and said second nucleic acid form a duplex, and wherein a plurality of said first nucleic acid is provided such that said first nucleic acid molecule is in concentration excess compared to said duplex.

- 74. The method of Claim 73, wherein said cleaving said cleavage structure comprises cleaving said first nucleic acid molecule to generate non-target cleavage product.
- 75. The method of Claim 74, wherein said non-target cleavage product from said first nucleic acid molecule is generated in concentration excess compared to said duplex.
- 76. The method of Claim 26, further comprising providing a third nucleic acid molecule complementary to a third portion of said target nucleic acid upstream of said first portion of said first target nucleic acid, wherein said cleavage structure comprises said third nucleic acid molecule.

FORM PTO-1449 (Modified) JAN 2 2 2003

U.S. Department of Commerce Patent and Trademark Office

Attorney Docket No.: FORS-06638

Serial No.: 09/982,667

INFORMATION DISCLOSURES TATEMENT BY APPLICANT (USE 10 PROPERTY OF THE PROPERTY

Applicant: James R. PRUDENT et al.

Filing Date: 10/18/01

Group Art Unit:

(37 CFR § 1.98(b))

U.S. PATENT DOCUMENTS

	]			U.S. PATENT DOCUMENTS	<del></del>		<u> </u>
Examiner Initials	Cite No.	Serial / Patent Number	Issue Date	Applicant / Patentee	Class	Subclass	Filing Date
	1	6,001,567	12/14/99	Brow et al.	435	6	07/12/96
	2	5,994,069	11/30/99	Hall et al.	435	6	03/24/97
	3	5,985,557	11/16/99	Prudent <i>et al</i> .	435	6	11/26/96
	4	5,888,780	03/30/99	Dahlberg et al.	435	91.53	02/19/97
	5	5,882,867	03/16/99	Ullman et al.	435	6	07/07/95
	6	5,874,283	02/23/99	Harrington et al.	435	252	05/30/95
•	7	5,846,717	12/08/98	Brow et al.	435	6	01/24/97
	8	5,843,669	12/01/98	Kaiser et al.	435	6	11/29/96
	9	5,843,654	12/01/98	Heisler et al.	435	6	07/07/95
	10	5,837,450	11/17/98	Dahlberg et al.	435	6	06/06/95
	11	5,830,664	11/03/98	Rosemeyer et al.	435	6	07/11/95
	12	5,795,763	08/18/98	Dahlberg et al.	435	194	06/06/95
	13	5,792,614	08/11/98	Western et al.	435	6	08/02/96
•	14	5,783,392	07/21/98	Seibl et al.	435	6	11/22/95
	15	5,719,028	02/17/98	Dahlberg et al.	435	6	02/06/97
•	16	5,698,400	12/16/97	Cotton et al.	435	6	09/16/96
_	17	5,691,142	11/25/97	Dahlberg et al.	435	6	06/06/96
	18	5,614,402	03/25/97	Dahlberg et al.	435	199	06/06/94
	19	5,601,976	02/11/97	Yamane et al.	435	6	09/16/92
	20	5, 545,729	08/13/96	Goodchild et al.	536	24.5	12/22/94
	21	5,541,311	07/30/96	Dahlberg et al.	536	23.7	06/04/93
	22	5,494,810	02/27/96	Barany et al.	435	91.52	11/22/94
	23	5,487,972	01/30/96	Geland et al.	435/6	435/91.2	01/05/93
	24	5,427,930	06/27/95	Birkenmeyer et al.	435	91/52	06/28/91
	25	5,422,253	06/06/95	Dahlberg et al.	435	91.53	12/07/92
	26	5,407,795	04/18/95	Kolberg et al.	435	5	10/15/93
	27	5,403,711	04/04/95	Walder et al	435	6	07/06/93
	28	5,660,988	08/26/97	Duck et al.	435/6	536/24.3	6/7/95
	29	5,380,833	06/10/95	Urdea	536	22.1	12/13/91
	30	5,210,015	05/11/93	Gelfand et al.	435	6	05/11/93
				A CONTRACTOR OF THE CONTRACTOR			
miner			1	Date Considered		L L	

Examiner:

Date Considered:

EXAMINER:

Initial citation considered. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

Translation

FORM PTO-1 (Modified)

U.S. Department of Commerce Patent and Trademark Office

Attorney Docket No.: FORS-06638

Serial No.: 09/982,667

SURE STATEMENT BY APPLICANT Several Sheets If Necessary)

Applicant: James R. PRUDENT et al.

(37 CFR § 1.	98(b))			Filing Date: 10/18/01		Group Art Un	it:	
U.S. PATENT DOCUMENTS								
Examiner Initials	Cite No.	Serial / Patent Number	Issue Date	Applicant / Patentee	Class	Subclass	Filing Date	
	T				1	1	1	

Examiner Initials	Cite No.	Serial / Patent Number	Issue Date	Applicant / Patentee	Class	Subclass	Filing Date
	31	5,144,019	09/01/92	Rossi	536	27	06/21/89
	32	5,118,605	06/02/92	Urdea	435	6	09/29/88
	33	5,108,892	04/28/92	Burke et al.	435	6	08/03/89
	34	5,030,557	07/09/91	Hogan et al.	435	6	11/24/87
	35	5,011,769	04/30/91	Duck et al.	435	6	04/29/88
	36	4,876,187	10/24/89	Duck et al.	435	6	12/05/85
	37	4,818,680	04/04/89	Collins et al.	435	6	12/18/85
	38	4,775,619	10/04/88	Urdea	435	6	10/16/84
•	39	4,683,202	07/28/87	Mullis	435	91	10/25/85
	40	4,683,195	07/28/87	Mullis et al.	435	6	02/07/86
	41	4,683,194	07/28/87	Saiki <i>et al</i> .	435/6	935/78	03/28/85
	42	4,518,526	05/21/85	Olson	260	112	06/01/84
	43	4,512,922	04/23/85	Jones et al.	260	112	06/01/84
•	44	4,511,503	04/16/85	Olson et al.	260	112	06/01/84
	45	4,511,502	04/16/85	Builder et al.	260	112	06/01/84

# FOREIGN PATENTS OR PUBLISHED FOREIGN PATENT APPLICATIONS

1	Document Number	Publication Date	Country / Patent Office	Class	Subclass	Yes	No
46	90/01069	02/08/90	PCT	C12Q	1/68		
47	92/06200	04/16/92	PCT	C12N	15/54		
48	91/09950	07/11/91	PCT	C12N	15/54		
 49	90/15157	12/13/90	PCT	C12Q	1/68		
50	96/40999	12/19/96	PCT	C12Q	C10P 19/34		
51	94/29482	12/22/94	PCT	C12Q 1/68	C12P 19/34		
52	95/14106	05/26/95	PCT	C12Q	1/68		
 53	92/02638	02/20/92	PCT	C12Q 1/68	1/70		
 54	89/09284	10/05/89	PCT	C12Q	1/68		
55	96/20287	07/04/96	PCT	C12Q 1/68	1/44		
·56	0 411 186 A1	02/06/91	European Patent Application	C12Q	1/68		
57	0 482 714 A1	10/22/91	European Patent Application	C12Q	1/68		
			*				

FORM PTO-1440 JAN 2 2 2003 (Modified).

U.S. Department of Commerce Patent and Trademark Office

Attorney Docket No.: FORS-06638

Serial No.: 09/982,667

INFORMATION DISCLOSURE STATEMENT BY APPLICANT Applicant: James R. PRUDENT et al. Group Art Unit: Filing Date: 10/18/01 (37 CFR § 1.98(b)) OTHER DOCUMENTS (Including Author, Title, Date, Relevant Pages, Place of Publication) Abrams et al., "Comprehensive Detection of Single Base Changes in Human Genomic DNA Using Denaturing Gradient Gel Electrophoresis 58 and a GC Clamp," Genomics 7:463-475 (1990) Akhmetzjanov and Vakhitov, "Molecular cloning and nucleotide sequence of the DNA polymerase gene from Thermus flavus," Nucl. Acids 59 Res. 20:5839 (1992) Altamirano et al., "Identification of Hepatitis C Virus Genotypes among Hospitalized Patients in British Columbia, Canada," J. Infect. Dis. 60 171:1034-1038 (1995). Anderson and Young, "Quantitative Filter Hybridization", in Nucleic Acid Hybridization, Eds Hames & Higgins, IRL Press, Washington, 61 DC, pp. 73-111 (1985) Electrophoresis, 2nd Edition, ed. Anthony T. Andrews, Clarendon Press, New York, New York (1986), pp. 153-154 62 Antao et al. "A thermodynamic study of unusually stable RNA and DNA hairpins," Nucl. Acids Res. 19:5901-5905 (1991) 63 Bambara et al., "Enzymes and Reactions at the Eukaryotic DNA Replication Fork," J. Biol. Chem. 272:4647-4650 (1997) 64 Barany, "Genetic disease detection and DNA amplification using cloned thermostable ligase," Proc. Natl. Acad. Sci., 88:189-193 (1991) 65 Barany, "The Ligase Chain Reaction in a PCR World," PCR Methods and Applic., 1:5-16 (1991) 66 Bardwell et al., "Specific Cleavage of Model Recombination and Repair Intermediates by the Yeast Rad1-Rad10 DNA Endonuclease," Science 265:2082-2085 (1994) 67 Barnes et al., "Mechanism of Tracking and Cleavage of Adduct-damaged DNA Substrates by the Mammalian 5'- to 68 3'Exonuclease/Endonuclease RAD2 Homologue 1 or Flap Endonuclease 1", J. Biol. Chem. 271:29624-29632 (1996) Bergseid et al., "A High Fidelity Thermostable DNA Polymerase Isolated from Pyrococcus Furiosus," Strategies 4:34-35 (1991) 69 Bhagwat et al., "The 5'-Exonuclease Activity of Bacteriophage T4 RNase H is Stimulated by the T4 Gene 32 Single-stranded DNA-binding 70 Protein, but Its Flap Endonuclease Is Inhibited," J. Biol. Chem. 272:28523-28530 (1997); Bonch-Osmolovskaya, et al., Microbiology (Engl. Transl. of Mikrobiologiya) 57:78-85 (1988) 71 Brutlag et al., "An Active Fragment of DNA Polymerase Produced By Proteolytic Cleavage," Biochem. Biophys. Res. Commun. 37:982-989 72 Brow et al., "Differentiation of Bacterial 16S rRNA Genes and Intergenic Regions and Mycobacterium tuberculosis katG Genes by 73 Structure-Specific Endonuclease Cleavage," J. of Clin. Micro. 34:3129-3137 (1996) Carballeira et al., "Purification of a Thermostable DNA Polymerase from Thermus thermophilus HB8, Useful in the Polymerase Chain 74 Reaction," Biotechniques 9:276-281 (1990) Ceska et al., "A helical arch allowing single-stranded DNA to thread through T5 5'-exonuclease," Nature 382:90-93 (1996) 75 Ceska et al., "Structure-specific DNA cleavage by 5' nucleases," TIPS 23 (1998) 76 Copley and Boot, "Exonuclease Cycling Assay: An Amplified Assay for the Detection of Specific DNA Sequences," BioTechniques 13:888-77 89i (1992) Cuthbert, "Hepatitis C:Progress and Problems," Clin. Microbiol. Rev. 7:505-532 (1994) 78 DeMott et al., "Human RAD2 Homolog 1 5'-3'-Exo/Endonuclease Can Efficiently Excise a Displaced DNA Fragment Containing a 5'-79 Terminal Abasic Lesion by Endonuclease Activity," J. Biol. Chem. 271:30068-30076 (1996) Doty et al., "Strand Separation and Specific Recombination in Deoxyribonucleic Acids: Physical Chemical Studies," Proc. Natl. Acad. Sci. 80 USA 46:461-476 (1960) Duck et al., "Probe Amplifier System Based on Chimeric Cycling Oligonucleotides," BioTech., 9:142-147 (1990) 81 Dunn et al., "Complete Nucleotide Sequence of Bacteriophage T7 DNA and the Locations of T7 Genetic Elements," J. Mol. Biol. 166:477-82 535 (1983) Date Considered: Examiner: Initial citation considered. Draw line through citation if not in conformance and not considered. Include copy of this form EXAMINER: with next communication to applicant.

FORM PTO-1 (Modified), INFORMATION DISCOSURE STATEMENT BY APPLICANT

U.S. Department of Commerce Patent and Trademark Office

Attorney Docket No.: FORS-06638

Serial No.: 09/982,667

with next communication to applicant.

INFOR	MATIG	N DISCOSURE STATEMENT BY APPLICANT	Applicant: James R. PRUDENT et al.	· · · · · · · · · · · · · · · · · · ·				
7 CFR § 1.98(	(b))	Expension sinces in recessary)	Filing Date: 10/18/01	Group Art Unit:				
		OTHER DOCUMENTS (Including Author, Title, D	ate, Relevant Pages, Place of Publication)					
	83	Engelke, "Purification of Thermus Aquaticus DNA Polymerase Expressed in Escherichia coli," Anal. Biochem 191:396-400 (1990)						
	84	Eom et al., "Structure of Taq polymerase with DNA at the polymerase active site," Nature 382:278-282 (1996)						
	85	Erlich et al., "Recent Advances in the Polymerase Chain Reac	tion," Science 252:1643-1651 (1991)					
	Fahy et al., "Self-sustained Sequence Replication (3SR): An Isothermal Transcription-based Amplification System Alter PCR Meth. Appl., 1:25-33 (1991)							
	87	Garforth et al., "Structure-specific DNA binding by bacterioph	nage T5 5'→3' exonuclease," Nucleic Acid.	s Res. 25:3801-3807 (1997)				
	88	Gelfand, PCR Technology - Principles and Applications for D	NA Amplification (H.A. Erlich, Ed.), Stoc	kton Press, New York, p. 19 (1989)				
	89	Guatelli et al., "Isothermal, in vitro amplification of nucleic ac Natl. Acad. Sci., 87:1874-1878 (1990) with an erratum at Production	cids by a multienzyme reaction modeled affic. Natl. Acad. Sci., 87:7797 (1990)	ter retroviral replication," Proc.				
•	90	Harrington et al., "DNA Structural Elements Required for FEN	N-1 Binding," J. Biol. Chem. 270:4503-450	08 (1995)				
	91	Harrington et al., "The characterization of a mammalian DNA	sturcture-specific endonuclease," EMBO J	ourn. 13:1235-1246 (1994)				
•	92	Harrington and Lieber, "Functional domains within FEN-1 and nucleotide excision repair," Genes and Develop. 8:1344-1355	d RAD2 define a family of structure-specif (1994)	ic endonucleases: implications for				
	93	Hayashi, "PCR-SSCP: A Simple and Sensitive Method for Detection of Mutations in the Genomic DNA," PCR Meth. Appl., 1:34-38, (1991)						
	94	Higuchi, R., (Ehrlich, H.A. (Ed.)), PCR Technology: Principle	es and Applications for DNA Amplification	, Stockton Press, pp. 61-70 (199				
	95	Hiraro et al. "Most compact hairpin-turn structure exerted by structure resistant to nucleases and heat," Nuc. Acids Res. 22:	a short DNA fragment, d(GCGAAGC) in s 576-582 (1994)	olution: an extraordinarily stable				
•	96	Holland et al., "Detection of specific polymerase chain reaction DNA polymerase," Proc. Natl. Acad. Sci. USA 88:7276-7280	on product by utilizing the 5'-3' exonucleas (1991)	e activity of Thermus aquaticus				
	97	Hosfield et al., "Structure of the DNA Repair and Replication FEN-1 Activity," Cell 95:135-146 (1996)	Endonuclease and Exonuclease FEN-1: C	Coupling DNA and PCNA Binding				
	98	Hosfield et al., "Newly Discovered Archaebacterial Flap Endo and Catalysis Resembling Human Flap Endonuclease-1," J. Bi	onucleases Show a Structure-Specific Mechaiol. Chem. 273:27154-17161	anism for DNA Substrate Binding				
	99	Huang et al., "Role of Calf RTH-1 Nuclease in Removal of 5' 9277 (1996)	'-Ribonucleotides during Okazaki Frament	Processing," Biochemistry 35:926				
	100	Hwang et al., "The crystal structure of flap endonuclease-1 from Methanococcus jannaschii," Nature Structural Biology 5:707-713 (1998);						
	101	Inchauspe et al., "Use of Conserved Sequences from Hepatitis C Virus for the Detection of Viral RNA in Infected Sera by Polymerase Chain Reaction," Hepatology 14:595-600 (1991)						
	102	Ito et al., "Compilation and alignment of DNA polymerase see	quences," Nucl. Acids Res. 19:4045-4057 (	1991)				
	103	Jacob and Monod, "On the Regulation of Gene Activity," Col	d Springs Harbor Symposium on Quantitat	ive Biol. XXVI:193-211 (1961)				
	104	Johnson et al., "Requirement of the Yeast RTH1 5' to 3' Exonuclease for the Stability of Simple Repetitive DNA," Science 269:238-240 (1995)						
	105	Kaledin et al., "Isolation and Properties of DNA Polymerase From the Extremely Thermophilic Bacterium Thermus flavus," Biokhimiya 46(9):1576-1584 (1981)						
	106	Kim et al., "Crystal structure of Thermus aquaticus DNA poly	ymerase," Nature 376:612-616 (1995)					
	107	Kornberg, DNA Replication, W.H. Freeman and Co., San Fra	incisco, pp. 127-139 (1980)					
	108	Kotler et al., "DNA sequencing: Modular primers assembled 90:4241-4245 (1993)	from a library of hexamers or pentamers,"	Proc. Natl. Acad. Sci. USA				
	109	Kwoh et al., "Transcription-based amplification system and desandwich hybridization format," Proc. Natl. Acad. Sci., 86:11	etection of amplified human immunodeficie 73-1177 (1989)	ency virus type 1 with a bead-base				
	110	Kwok et al., "Effects of primer-template mismatches on the p-studies," Nucl. Acids Res., 18:999-1005 (1990)	olymerase chain reaction: Human immuno	deficiency virus type 1 model				
aminer:			Date Considered:					



FORM PTO-1449 (Modified). U.S. Department of Commerce Patent and Trademark Office

Attorney Docket No.: FORS-06638

Serial No.: 09/982,667

INFORMATION DISCUSSIVE STATEMENT BY APPLICANT (Use Several Sheets If Necessary)			Applicant: James R. PRUDENT et al.					
(37 CFR § 1.9	8(b))	(OSC OCTORAL OLICOS II INCCOSSALJ)	Filing Date: 10/18/01	Group Art Unit:				
·		OTHER DOCUMENTS (Including Author, Title, D	e, Date, Relevant Pages, Place of Publication)					
	Landegren, "Molecular mechanics of nucleic acid sequence amplification," Trends in Genetics 9:199-204 (1993)							
	112	Lawyer et al., "Isolation, Characterization, and Expression in Escherichia coli of the DNA Polymerase Gene from Thermus aquaticus," J. Biol. Chem. 264:6427-6437 (1989)						
	113	Leirmo et al., "Replacement of Potassium Chloride by Potassi Biochem. 26:2095-2101 (1987)	ium Glutamate Dramatically Enhances Prot	ein-DNA Interactions in Vitro,"				
	114	Levine, "The Tumor Suppressor Genes," Annu. Rev. Biochen	n. 62:623 (1993)					
	ulation of FEN-1 by Proliferating							
	116	Lieber, "The FEN-1 family of structure-specific nucleases in (1997)	eukaryotic DNA replication, recombination	and repair," BioEssays 19:233-240				
r	117	Lindahl, et al., "Deoxyribonuclease IV: A New Exonuclease	From Mammalian Tissues," Proc. N.A.S. (	52:597-603 (1968)				
	118	Lindahl and Karlström, "Heat-Induced Depyrimidination of D	eoxyribonucleic Acid in Neutral Solution,"	Biochem. 12:5151-5154 (1973)				
•	119	Longley et al. "Characterization of the 5' to 3' exonuclease at 7322 (1990)	ssociated with Thermus aquaticus DNA po	lymerase," Nucl. Acids Res. 18:731				
	120	Lundquist, et al., "Transient Generation of Displaced Single-S	Stranded DNA during Nick Translation," Co	ell 31:53-60 (1982)				
	121	Lyamichev et al."Structure-Specific Endonucleolytic Cleavage (1993)	e of Nucleic Acids by Eubacterial DNA Po	lymerases," Science 260:778-783				
	122	Marmur and Lane, "Strand Separation and Specific Recombination in Deoxyribonucleic acids: Biological Studies," <i>Proc. Natl. Acad. Sci. USA</i> 46:453-461 (1960)						
•	123	Mathur et al., "The DNA polymerase gene from the hyperthermophilic marine archaebacterium Pyrococcus furiosus, shows sequence homology with α-like DNA polymerases," Nucl. Acids Res. 19:6952 (1991)						
٧	124	Milligan and Ublenbeck, "Synthesis of Small RNAs Using T7 RNA Polymerase," Methods Enzymol. 180:51 (1989)						
	125	Milligan et al., "Oligoribonucleotide synthesis using T7 RNA polymerase and synthetic DNA templates," Nucl. Acids. Res. 15(21): 8783-8789 (1987)						
	126	Mullis, "The Polymerase Chain Reaction in an Anemic Mode: How to Avoid Cold Oligodeoxyribonuclear Fusion," PCR Methods Applic., 1:1-4 (1991)						
	127	Mullis and Faloona, "Specific Synthesis of DNA in Vitro via a Polymerase-Catalyzed Chain Reaction," Methods in Enzymology 155:335-350 (1987)						
	128	Murante et al., "Calf 5' to 3' Exo/Endonuclease Must Slide from a 5' End of the Substrate to Perform Structure-specific Cleavage," J. Biol. Chem. 270:30377-30383 (1995)						
	129	Murante et al., "The Calf 5'- to 3'-Exonuclease Is Also an E the Point of Cleavage," J. Biol. Chem. 269:1191-1196 (1994)	ndonuclease with Both Activities Dependen	nt on Primers Annealed Upstream of				
Ò	130	Murray et al., "Structural and Functional Conversation of the Required for Chromosome Segregation and Recovery from D						
	131	Myers et al., "Reverse Transcription and DNA amplification by	y a Thermus thermophilus DNA Polymera	se," Biochem. 30:7661-7666 (1991				
	132	Nielsen PE et al., "Peptide nucleic acids (PNAs): Potential an	ti-sense and anti-gene agents," Anticancer	Drug Des. 8:53-63 (1993)				
	133	Nolan et al., "Kinetic Analysis of Human Flap Endonuclease-	1 by Flow Cytometry," Biochemistry 35:11	668-11677 (1996)				
	134	Nugent et al., "Characterization of the Apurinic Endonuclease	Activity of Drosophila Rrpl," Biochemistr	y 32:11445-11452 (1993)				
	135							
	136	Pontius and Berg, "Rapid renaturation of complementary DNA domains in enhancing the kinetics of molecular assembly produced in the complementary and produced in the complementary DNA domains and Berg, "Rapid renaturation of complementary DNA domains and Berg," and "Rapid renaturation of complementary DNA domains and Berg," and "Rapid renaturation of complementary DNA domains and Berg," and "Rapid renaturation of complementary DNA domains and Berg," and "Rapid renaturation of complementary DNA domains are complementary DNA domains and Berg," and "Rapid renaturation of complementary DNA domains are complementary DNA domains and Berg," and "Rapid renaturation of complementary DNA domains are complementary DNA domains and "Rapid renaturation" and "Rapid renaturatio						
Examiner:			Date Considered:					
EXAMINER:		tial citation considered. Draw line through citation if not in con	formance and not considered. Include cop	y of this form				
····	Wil	th next communication to applicant.	<u> </u>					

FORM PTO-14(S) (Modified).

U.S. Department of Commerce Patent and Trademark Office

Attorney Docket No.: FÖRS-06638

Serial No.: 09/982,667

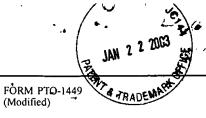
INFORMA HORADE COSURE STATEMENT BY APPLICANT (Use Several Sheets If Necessary)

Applicant: James R. PRUDENT et al.

	(Use Several Sheets If Necessary)					
(37 CFR § 1.98(b))	(00000000000000000000000000000000000000	Filing Date: 10/18/01	Group Art Unit:			
<u> </u>	OTHER DOCUMENTS (Including Author, Title, D	Pate, Relevant Pages, Place of Publication)				
137	Rao et al., "Methanococcus jannaschii Flap Endonuclease: E. 180:5406-5412 (1998);	xpression, Purification, and Substrate Requ	irements," J. of Bacteriology			
138	Reagan et al., "Characterization of a Mutant Strain of Sacchar of the RAD2 Nucleotide Excision Repair Gene," J. of Bacterio	romyces cerevisiae with a Deletion of the Rology 177:364-371 (1995);	2AD27 Gene, a Structural Homolog			
139	Saiki et al., "Primer-Directed Enzymatic Amplification of DN	A with a Thermostable DNA Polymerase,"	Science 239:487-491 (1988);			
140	Sambrook et al., Molecular Cloning. A Laboratory Manual, (1989);	Cold Spring Harbor Laboratory Press, Cold	Spring Harbor, pp. 1.63-1.69			
141	Setlow and Kornberg, "Deoxyribonucleic Acid Polymerase: To	wo Distinct Enzymes in One Polypeptide,"	J. Biol. Chem. 247:232-240 (1972)			
142	Siegal et al., "A 5' to 3' exonuclease functionally interacts wi	th calf DNA polymerase E," Proc. Natl. Ac	ad. Sci. USA 89:9377-9381 (1992)			
143	Shen et al., "Flap endonuclease homologs in archaebacteria ex	xist as independent proteins," TIBS 23 (199	8);			
144	Shen et al., "Essential Amino Acids for Substrate Binding and (1996)	1 Catalysis of Human Flap Endonuclease 1	" J. of Biol. Chem. 271:9173-9176			
145	Smith et al., "Novel Method of Detecting Single Base Substitu Genomics 3:217-223 (1988);	utions in RNA Molecules by Differential M	lelting Behavior in Solution,"			
146	Sommers et al., "Conditional Lethality of Null Mutations in R Exonuclease Required for Lagging Strand DNA Synthesis in I					
147	Stark, "Multicopy expression vectors carrying the <i>lac</i> represso 5:255-267 (1987);	or gene for regulated high-level expression	of genes in Escherichia coli," Gen			
148	Studier and Moffatt, "Use of Bacteriophage T7 RNA Polymers 189:113-130 (1986);	Studier and Moffatt, "Use of Bacteriophage T7 RNA Polymerase to Direct Selective High-level Expression of Cloned Genes," J. Ma 189:113-130 (1986);				
149	Tindall and Kunkel, "Fidelity of DNA by the Thermus aquation	cus DNA Polymerase," Biochem. 27:6008-0	6013 (1988);			
150	Turchi et al., "Enzymatic completion of mammalian lagging-s	trand DNa replication," Proc. Natl. Acad. S	Sci. USA 91:9803-9807 (1994);			
151	Uhlenbeck, "A small catalytic oligoribonucleotide," Nature 32	8:596-600 (1987);				
152	Urdea et al., "A novel method for the rapid detection of speciradioactivity; application to the analysis if hepatitis B virus in		l samples without blotting or			
153	Wu and Wallace, "The Ligation Amplification Reaction (LAR Template-Dependent Ligation," <i>Genomics</i> 4:560-569 (1989);	2) - Amplification of Specific DNA Sequence	ces Using Sequential Rounds of			
154	Wu et al., "Processing of branched DNA intermediates by a co (1996);	omplex of human FEN-1 and PCNA," Nuc	leic Acids Research 24:2036-2043			
155	Xu et al., "Biochemical and Mutational Studies of the 5'-3' Ex 302 (1997);	xonuclease of DNA Polymerase 1 of Esche	erichia coli," J. Mol. Biol. 268:284			
156	Zwickl et al., "Glyceraldehyde-3-Phosphate Dehydrogenase fro Characterization of the Enzyme, Cloning and Sequencing of the					
157	Hiraoka et al., "Sequence of human FEN-1, a structure specifiand human," Genomics 25:220-225 (1995);	ic endonuclease, and chromosomal localiza	tion of the gene (FEN1) in mouse			
158	Augustyns et al., "Hybridization specificity, enzymatic activity beta-D-erythro-hexopyranosyl nucleosides," Nucleic Acids Res.		acleotides containing 2,4-dideoxy-			
159	Agrawal et al., "Modified oligonucleotides as therapeutic and	diagnostic agents," Current Opinion in Bio	technology, 6:12-19 (1995);			
160	Corey, "4800-fold Acceleration of Hybridization of Chemically (1995);	y Modified Oligonucleotides," J. of the Am	er. Chem. Soc. 117:9373-9374			
161	Cotton, "Current methods of mutation detection," Mutation Re-	search 285:125-144 (1993);				
162	Schmidt et al., "The use of oligonucleotide probes containing from Escherichia coli," Biochimica et Biophysica Acta. 1130:4		ific cleavage of RNA by RNaseH			
		T				
xaminer:		Date Considered:				

EXAMINER:

Initial citation considered. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.



U.S. Department of Commerce Patent and Trademark Office

Attorney Docket No.: FORS-06638

Applicant: James R. PRUDENT et al.

Serial No.: 09/982,667

INFORMATION DISCLOSURE STATEMENT BY APPLICANT (Use Several Sheets If Necessary)

Group Art Unit

37 CFR § 1.98(b))	Filing Date: 10/18	/01	Group Art Unit:				
	OTHER DOCUMENTS (Including Author, Title, Date, Relevant Pages,	Place of Publication)					
163	Lee et al., "Allelic discrimination by nick-translation PCR with fluorogenic probes," Nucleic Acids Res. 21(16):3761-3766 (1993)						
164	Livak et al., "Oligonucleotides With Fluorescent Dyes at Opposite Ends Provide a Quenched Probe System, Useful for D Product and Nucleic Acid Hybridization," PCR Methods and Appln. 4:357-362 (1995)						
165	Gamper et al., "Solution Hybridization of Crosslinkable DNA Oligonucleotides to (1987)	Bacteriophage M13 DI	NA," <i>J. Mol. Biol.</i> 197:349-362				
166	Lima et al., "Implication of RNA Structure on Antisense Oligonucleotide Hybridiz	ation Kinetics," Bioche	mistry 31:12055-12061 (1992)				
167	Sigman et al., "Chemical Nucleases," Chem. Rev. 93:2295 (1993)	<del></del>					
168	Youil et al., "Screening for Mutations by Enzyme Mismatch Cleavage with T4 En (1995)	donuclease VII," <i>Proc.</i>	Natl. Acad. Sci. USA 92:87-91				
169	Abramson et al., "Characterization of the 5'-3' Exonuclease Activity of Thermus A	quaticus DNA Polyme	erase," FASEB J. 5(4) 386 (1991)				
170	Roychoudhury and Wu, "Novel Properties of Escherichia coli Exonuclease III," J.	Biol. Chem. 252:4786	-4789 (1977)				
ı'							
•							
•							
	ı						
	•						
xaminer:	Date Considered:						
	nitial citation considered. Draw line through citation if not in conformance and not convith next communication to applicant.	nsidered. Include copy	of this form				